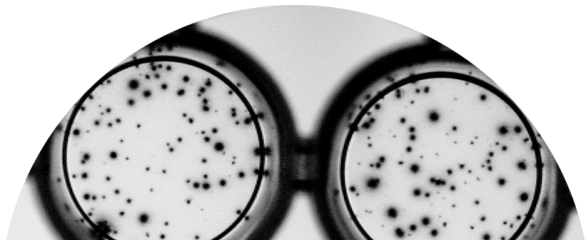
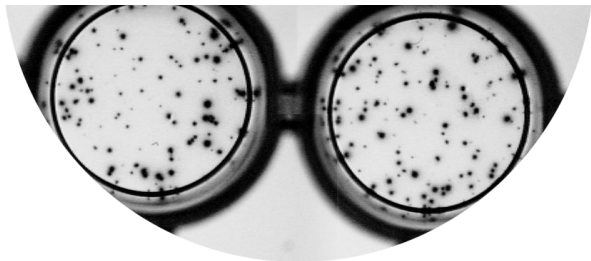


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# Instruction Manual T cell ELISPOT kit



Silver staining procedure  
on transparent plates



2-plate and 5-plate format

*For research use only.*  
Not for use in diagnostic or therapeutic procedures.





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# Abbreviations

APC	Antigen-Presenting Cell
CTL	Cytotoxic T Lymphocyte
CD	Cluster of Differentiation
CNS	Central Nervous System
ConA	Concanavalin A
ELISA	Enzyme-Linked Immunoassay
ELISPOT	Enzyme-Linked ImmunoSPOT
FCS	Fetal Calf Serum
GABA	Gold-labeled Anti-Biotin Antibodies
ICE	Influenza A, Cytomegalovirus, Epstein-Barr virus
IFN	Interferon
IL	Interleukin
LPA	Lymphocyte Proliferation Assay
MHC	Major Histocompatibility Complex
min	minute(s)
PBS	Phosphate Buffered Saline
PBS-I	Sterile and Pyrogen-free PBS
PBMC	Peripheral Blood Mononuclear Cell
PHA	Phytohaemagglutinin
PMA	Phorbol 12-Myristate 13-Acetate
RT	Room Temperature (temperature between 20 °C and 26 °C)
RT-PCR	Real Time Polymerase Chain Reaction
sec	seconds

# Catalogue numbers T cell ELISPOT kits

This manual applies to the following T cell ELISPOT kits

Analyte	Human	Old World Monkey	Mouse	Rat
IFN- $\gamma$	CT230-T2 (2-plate) CT230-T5 (5-plate)	CT121-T2 (2-plate) CT121-T5 (5-plate) CT126-T2 (2-plate) CT126-T5 (5-plate)	CT317-T2 (2-plate) CT317-T5 (5-plate)	CT079-T2 (2-plate) CT079-T5 (5-plate)
IL-2	CT231-T2 (2-plate) CT231-T5 (5-plate)	CT127-T2 (2-plate) CT127-T5 (5-plate)		
IL-4	CT232-T2 (2-plate) CT232-T5 (5-plate)	CT128-T2 (2-plate) CT128-T5 (5-plate)	CT319-T2 (2-plate) CT319-T5 (5-plate)	CT081-T2 (2-plate) CT081-T5 (5-plate)
IL-5	CT233-T2 (2-plate) CT233-T5 (5-plate)	CT129-T2 (2-plate) CT129-T5 (5-plate)	CT321-T2 (2-plate) CT321-T5 (5-plate)	
IL-6	CT234-T2 (2-plate) CT234-T5 (5-plate)	CT130-T2 (2-plate) CT130-T5 (5-plate)	CT436-T2 (2-plate) CT436-T5 (5-plate)	
IL-10	CT235-T2 (2-plate) CT235-T5 (5-plate)	CT131-T2 (2-plate) CT131-T5 (5-plate)	CT320-T2 (2-plate) CT320-T5 (5-plate)	
IL-12/23p40		CT135-T2 (2-plate) CT135-T5 (5-plate)		
IL-12p70	CT240-T2 (2-plate) CT240-T5 (5-plate)			
IL-13	CT236-T2 (2-plate) CT236-T5 (5-plate)	CT132-T2 (2-plate) CT132-T5 (5-plate)		
IL-17A	CT416-T2 (2-plate) CT416-T5 (5-plate)	CT401-T2 (2-plate) CT401-T5 (5-plate)		
IL-17F	CT418-T2 (2-plate) CT418-T5 (5-plate)	CT403-T2 (2-plate) CT403-T5 (5-plate)		
Perforin	CT681-T2 (2-plate) CT681-T5 (5-plate)	CT136-T2 (2-plate) CT136-T5 (5-plate)		
TNF- $\alpha$	CT237-T2 (2-plate) CT237-T5 (5-plate)	CT133-T2 (2-plate) CT133-T5 (5-plate)	CT322-T2 (2-plate) CT322-T5 (5-plate)	

NOTE: the accompanying ‘Typical data’ and ‘Addendum T cell ELISPOT assay’, which contains guidelines and troubleshooting for ELISPOT analyses are available on our website ([www.ucytech.com/manuals](http://www.ucytech.com/manuals)).

# Introduction

The ELISPOT assay is one of the most sensitive tests to monitor *ex-vivo* cellular immune responses at the single cell level. The assay can accurately detect secreted proteins, such as cytokines, released by T cells in response to an antigen. The cell suspensions, used in the test, can originate from blood (PBMCs), lymphoid, spleen, bone marrow or CNS tissue.

Classical T cell monitoring assays (e.g. LPA and CTL assay), measure CD4<sup>+</sup> or CD8<sup>+</sup> cell mediated immune responses. Both LPA and CTL assays have their drawbacks including the use of radioactivity, low throughput screening, decreased sensitivity in cryopreserved specimens and technical burden. RT-PCR analysis, to measure T cell responses can also be used. However, this assay detects mRNA instead of actually secreted protein.

The ELISPOT assay, not afflicted with these shortcomings, has proven to be more sensitive than an ELISA<sup>1</sup> or intracellular cytokine staining<sup>2,4</sup>. The high sensitivity is possible due to the plate-bound antibodies that directly capture the secreted proteins released by the cell before they dilute in the culture medium, are taken up by cells via cell-surface receptors or are degraded by proteases. This property enables the detection of very low frequencies of cytokine secreting cells (1/300,000) and also offers the possibility of high throughput screening.

The ELISPOT procedure, based on silver staining, makes use of affordably priced transparent polystyrene flat-bottomed plates. Each 5-plate kit is provided with 6 plates and the 2-plate kit with 2 plates. Spot staining is achieved with gold-labeled anti-biotin antibodies (GABA) using silver precipitation for spot visualization, a highly effective and sensitive method. Spots do not fade in time and ELISPOT plates can be reanalyzed several years later after being stored at room temperature.

Nowadays, our ELISPOT procedure with silver staining on transparent plates is widely used in different fields of biomedical research, including the diabetes field. For example, Arif *et al.* (2004)<sup>3</sup> developed the so-called UK-ELISPOT with our ELISPOT procedure, which has proven to have a significant discriminative ability for type 1 diabetes in blinded proficiency testing.<sup>4,6</sup>

## References

1. Tanguay and Killion (1994). Direct comparison of ELISPOT and ELISA-based assays for detection of individual cytokine-secreting cells. *Lymphokine Cytokine Res* 13: 259.
2. Carter and Swain (1997). Single cell analyses of cytokine production. *Curr Opin Immunol* 9: 177.
3. Arif *et al.* (2004). Autoreactive T cell responses show proinflammatory polarization in diabetes but a regulatory phenotype in health. *J Clin Invest* 113: 451-63.
4. Herold *et al.* (2009). Validity and reproducibility of measurement of islet autoreactivity by T-cell assays in subjects with early type 1 diabetes. *Diabetes* 58: 2588-95.
5. Gómez-Touriño *et al.* (2015). Characterization of the autoimmune response against the nerve tissue S100B in patients with type 1 diabetes. *Clin Exp Immunol* 180: 207-217.
6. Arif *et al.* (2014). Blood and islet phenotypes indicate immunological heterogeneity in type 1 diabetes. *Diabetes* 63(11): 3835-45.

Please find more references of studies using our ELISPOT kits on our website:  
[www.ucytech.com/ELISPOT](http://www.ucytech.com/ELISPOT) or [www.ucytech.com/references](http://www.ucytech.com/references).

# Brief description ELISPOT assay

U-CyTech ELISPOT kits are simple and sensitive immunoassays for the detection of protein secreting cells at the single cell level.

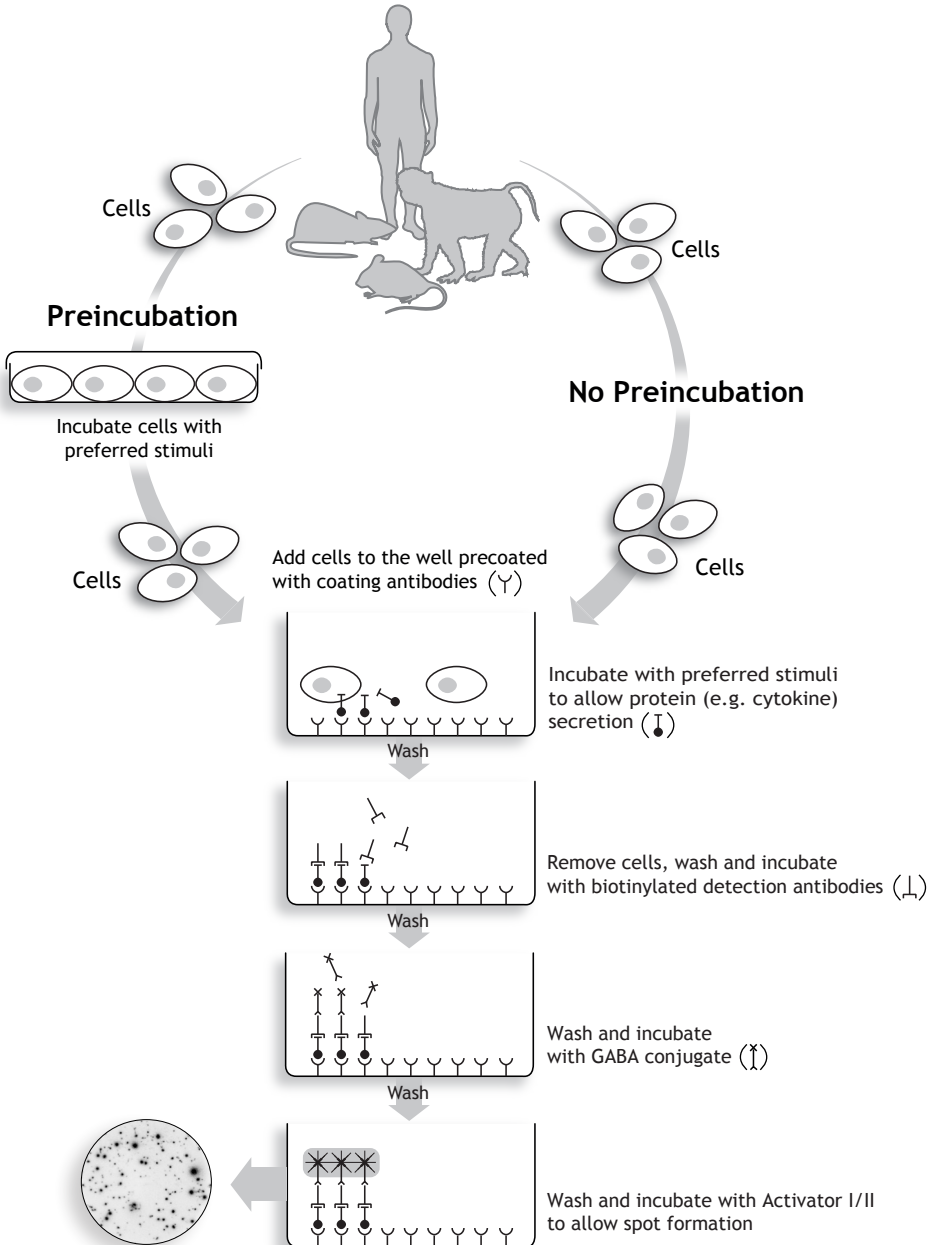
The ELISPOT procedure is illustrated in the “Flow diagram ELISPOT” on page 8. First cells are collected from a donor. Depending on the experimental set-up, the cells are either preincubated (see the next paragraph) or directly added to the wells of the ELISPOT plate that is coated with a high affinity antibody\*. The cell suspension is incubated in the presence of an antigen or polyclonal stimulus to trigger protein release from the cell. Subsequently, the cells are washed away and the antibody-bound proteins are detected with a combination of a biotinylated detection antibody and GABA conjugate. In the last step, a silver salt solution (Activator I/II) is added which allows silver to precipitate on the gold particles producing a spot that reveals the protein secretion site (footprints of individual cells).

Preincubation (24-48 hours) is required when full-length proteins or long peptides are used for re-stimulation. These antigens must first be internalized, processed and presented by antigen-presenting cells (APCs) via MHC class I/II molecules before they can stimulate protein (e.g. cytokine) release by T cells. Omitting this step leads to a significant lower frequency of spot forming cells. On the other hand, small peptides (8-12 amino acids) can directly be presented by APC to CD8<sup>+</sup> cells and consequently need no preincubation step. Read “Cell sample preparation” on page 14 for more information.

\* specific for a protein of interest

# Flow diagram ELISPOT

Sterile conditions



Non-sterile conditions



# Warnings and precautions

- This kit is designed for research use only, and not for use in diagnostic or therapeutic procedures.
- Please note that human and non-human primate blood components or other biological materials should be considered as potentially infectious and handled with the usual precautions under Bio-Hazard conditions. Follow universal precautions such as established by the US government agencies, Centers for Disease Control and Prevention and Occupational Safety and Health Administration, when handling and disposing of (potentially) infectious waste.

## Hazard information

Except for the Activator I+II solutions, the items in this kit are not classified as dangerous according to Regulation (EC) no. 1272/2008 and its amendments.

Activator I + II:



### **Warning:**

Activator I+II solutions are classified as dangerous according to Regulation (EC) no. 1272/2008 and its amendments: Serious eye damage (Category 1), Skin sensitization (Category 1) and Chronic aquatic toxicity (Category 1).

Hazard statement: May cause an allergic skin reaction (H317), Causes serious eye damage (H318), Very toxic to aquatic life with long lasting effects (H410).

The Activator I+II solutions should be kept away from acids and sources of ignition; use only non-sparking tools. Keep away from light, air and heat. In case of contact with skin, wash with soap and water and remove contaminated clothing and shoes. Upon ingestion rinse mouth (if person is conscious) and call physician immediately. Do not induce vomiting. In case of contact with eyes, irrigate with copious amounts of water for at least 15 min. Assure adequate flushing by separating the eyelids. Seek medical advice immediately.

Please find the Material and Safety Data Sheet on [www.ucytech.com/manuals](http://www.ucytech.com/manuals).

# Contents of the kit

Items	Quantity (2-plate format)	Quantity (5-plate format)	Storage conditions
Coating antibody*	1 vial	1 vial	4 °C
Biotinylated detection antibody*	1 vial	1 vial	4 °C
GABA conjugate*	1 vial	1 vial	4 °C
Activator I	4 ml	9.5 ml	4 °C in the dark
Activator II	4 ml	9.5 ml	4 °C in the dark
Blocking stock solution (10x)	4 ml	10 ml	4 °C
Dilution buffer T (10x)	3.5 ml	8 ml	4 °C
Tween-20	5 ml	5 ml	RT
96-well ELISPOT plate** with lid	2	6	RT
Adhesive cover slip	5	10	RT

\* Lyophilized

\*\* Transparent polystyrene flat-bottomed Nunc MaxiSorp plates.

# Storage and stability

## **Coating antibody and biotinylated detection antibody**

The vials with lyophilized coating antibody and biotinylated detection antibody can be safely stored at 4°C until the expiry date (indicated on the vials). After reconstitution, the antibodies are stable for at least 12 months at 4°C when kept sterile. However, it is recommended to divide the reconstituted antibody solutions into small aliquots for single use. These aliquots should be stored at  $\leq -20^{\circ}\text{C}$  (stable for at least two years).

## **GABA conjugate**

The vial with lyophilized GABA conjugate can be safely stored at 4°C until the expiry date (indicated on the vial). After reconstitution, it is strongly recommended to divide the solution into small aliquots for single use at  $\leq -20^{\circ}\text{C}$  (stable for at least two years).

## **Activators I and II**

The Activators I and II should be stored at 4°C and are stable until the expiry date (indicated on the vials)\*. It is strongly recommended to divide the solutions into small aliquots for single use. These aliquots should be stored at  $\leq -20^{\circ}\text{C}$  in the dark (stable for at least two years).

\*Avoid exposure to light and air and tightly close the vials after use.

## **Blocking stock solution (10x) and Dilution buffer T (10x)**

The vials with blocking stock solution and Dilution buffer T can be safely stored at 4°C until the expiry date (indicated on the vials). After opening these solutions are stable for at least 6 months when kept sterile.

## **Tween-20**

Tween-20 can safely be stored at RT and is stable until the expiry date (indicated on the vial).

# Materials and equipment (required but not provided)

- Tubes and containers/plates to prepare the solutions.
- Tissue culture plates for preincubation (optional).
- Sterile distilled water.
- PBS pH 7.4 (home-made). For washing purposes only.
- Sterile and pyrogen-free PBS pH 7.4 (PBS-I): Thermo Fisher Scientific cat. no. 10010 is recommended (Gibco®).
- Culture medium: RPMI-1640 supplemented with 2 mM L-Glutamine, 100 units/ml penicillin, 100 µg/ml streptomycin and 10% fetal calf serum (FCS).
  - RPMI-1640: Thermo Fisher Scientific cat. no. 52400 (Gibco®).
  - L-Glutamine: Thermo Fisher Scientific cat. no. 25030-081 (Gibco®; 200 mM).
  - Penicillin-Streptomycin: Thermo Fisher Scientific cat. no. 15140-122 (Gibco®, 100x).
  - Fetal bovine serum should be selected on low background staining: ThermoFisher Scientific cat. no. 16000 (Gibco®).

Culture medium AIM V® (Thermo Fisher Scientific cat. no. 31035-025) supplemented with 100 units/ml penicillin and 100 µg/ml streptomycin is an alternative culture medium for the procedure without a preincubation step.

- Cell stimuli, see “Cell sample preparation” on page 14 and [www.ucytech.com/ELISPOT-stimuli](http://www.ucytech.com/ELISPOT-stimuli).
- Pipetting devices.
- For washing: squirt (wash or squeeze) bottle with wide sprout or (automated) washing device, see Addendum\*.
- CO<sub>2</sub> incubator (37°C, 100% humidity, 5% CO<sub>2</sub>).
- 37°C incubator.
- A transmitted light microscope or an Immunospot image analyzer for spot counting.

# Preparation solutions and reagents

*Prepare reagents under sterile conditions (e.g. Laminar Flow Hood)*

## **Blocking buffer (1x)**

Dilute Blocking stock solution (10x) in PBS-I.

For one ELISPOT plate: 2 ml is gently and thoroughly mixed with 18 ml PBS-I

## **Dilution buffer T (1x)**

Dilute Dilution buffer T (10x) in PBS-I.

For one ELISPOT plate: 1.5 ml is gently and thoroughly mixed with 13.5 ml PBS-I

## **Coating antibody**

Reconstitute the lyophilized contents by injecting an appropriate volume (indicated on vial) of sterile distilled water. Mix the solution gently for approximately 15 seconds and allow the vial to stand for 5 minutes at RT.

For one ELISPOT plate (2-plate kit): 100 µl is gently and thoroughly mixed with 5 ml PBS-I

For one ELISPOT plate (5-plate kit): 50 µl is gently and thoroughly mixed with 5 ml PBS-I

## **Biotinylated detection antibody**

Reconstitute the lyophilized contents of the vial by injecting an appropriate volume (indicated on vial) of sterile distilled water. Mix the solution gently for approximately 15 seconds and allow it to stand for 5 min at RT.

For one ELISPOT plate: 100 µl is gently and thoroughly mixed with 10 ml Dilution buffer T (1x).

## **GABA conjugate**

Reconstitute the lyophilized contents of the vial by injecting an appropriate volume (indicated on vial) of sterile distilled water. Mix the solution gently for approximately 15 seconds and allow it to stand for 5 min at RT.

For one ELISPOT plate: 100 µl is gently and thoroughly mixed with 5 ml Dilution buffer T (1x).

## **Activator I and II**

Bring the two bottles of Activator to RT prior to use. Shake the bottles gently but intensively before mixing 1.8 ml of Activator I with 1.8 ml Activator II (for one ELISPOT plate). Use immediately thereafter.

## **PBS (for washing purposes only)**

5.4 mM  $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ ; 1.3 mM  $\text{KH}_2\text{PO}_4$ ; 150 mM NaCl; pH 7.4 (sterile).

For one ELISPOT plate: prepare 1 L PBS.

## **Wash buffer**

PBS containing 0.05% Tween-20.

For one ELISPOT plate: 0.5 ml Tween-20 is gently and thoroughly mixed with 1 L PBS.

# Cell sample preparation

Both fresh and cryopreserved cells can be used for ELISPOT analysis. Guidelines for specimen collection and handling are described in the “Addendum T cell ELISPOT assay”.

Optimal conditions for the generation of cells releasing cytokines or other effector molecules in heterogeneous cell populations should be determined in advance. This is advisable because different cell types, producing the same effector molecules, require different conditions for stimulation. For instance, the optimal conditions for the detection of IFN- $\gamma$  secreting CD8<sup>+</sup> T cells in PBMC preparations differ considerably from those for the detection of IFN- $\gamma$  secreting CD4<sup>+</sup> T cells<sup>1</sup>.

Moreover, the production of cytokines, such as TNF- $\alpha$ , IL-6 and IL-10, is not restricted to T cells and many spot-forming cells can also be attributable to activated monocytes/macrophages. Adherence of these last type of cells to the surface of an ELISPOT well may already be sufficient to trigger TNF- $\alpha$  and IL-6 release.

## Assay Controls

Before starting an ELISPOT experiment, proper assay controls need to be chosen, which is mainly dependent on the selected analyte, targeted cell type and experimental set-up.

### Positive controls

As positive controls both antigen-specific and polyclonal stimuli can be used to demonstrate that the cells are functional and the assay works well. Well-defined reagents such as the ICE peptide pool (a pool of synthetic peptides of common viral epitopes cat. no. CT387) and monoclonal antibodies (e.g. anti-CD3/CD28 cat. no. CT372), are often preferred since these reagents are proven stimuli. In addition, also vaccine proteins (e.g. tetanus toxoid, Hepatitis B proteins) can be used, depending on whether all study subjects have been vaccinated.

Polyclonal stimuli such as, ConA, PHA, PMA/ionomycin, can be used for many different cell types of various species. An overview of ELISPOT stimuli and the recommended concentration ranges can be found in our “Addendum T cell ELISPOT assay” and on page 17. In general, the optimal antigen concentration for antigen-specific stimulations varies between 0.5 and 10  $\mu$ g of protein or peptide/ml, but should be determined experimentally.

### Negative controls

To reveal any false positive signals, or spontaneously secreting cells, cells are also incubated without stimuli at the same cell concentration as the cells incubated with the specific antigen of interest. In addition, a limited number of wells may be used including all reagents, but without the addition of cells, to exclude the possibility of false positivity due to the reagents or media.

## References

1. Schmittel *et al.* (2001). Application of the IFN-gamma ELISPOT assay to quantify T cell responses against proteins. *J Immunol Methods* 247: 17

### **Cell culture conditions preincubation**

A 24-48 hours preincubation step at high cell density ( $> 10^6$ ) may be required when full-length proteins or long peptides are used for *in vitro* re-stimulation. These long antigens must first be internalized, processed and presented by APCs via MHC class I/II molecules before they can stimulate cytokine (or other effector molecule) release by T cells. The high number of cells enhances the probability of contact between stimulating and responding cells. Omitting this step leads in most cases to a significant lower frequency of spot forming cells.

For preincubation, suspend cells in culture medium with an appropriate stimulus at  $4 \times 10^6$  cells/ml in a tissue culture plate and incubate 24-48 hours ( $37^\circ\text{C}$  with 5%  $\text{CO}_2$  in a humidified atmosphere). Use a minimum of 1 ml/well in a 24-well plate, 0.5 ml/well in a 48-well plate or 100  $\mu\text{l}$ /well in a 96-well plate.

After preincubation, the non-adherent cells are collected and washed twice with fresh culture medium without stimuli or fetal calf serum. This will avoid the carryover of cytokines or other effector molecules produced during the preincubation step (two centrifugation/resuspension steps; 8 min, 200x g, RT). Thereafter cells are counted and suspended in culture medium with the same stimulus as used during preincubation at  $1\text{-}3 \times 10^5$  cells/well (antigen-specific responses). For polyclonal stimulation, the recommended cell concentration per well should be reduced to  $2 \times 10^2$  -  $10^5$  cells per well.

### **Stimulation with small peptides (8-12 amino acids)**

Small peptides can directly be presented by APCs to T cells. Consequently such peptides can be used in the ELISPOT assay without a preincubation step. For antigen-specific stimulation,  $1\text{-}3 \times 10^5$  cells per well is recommended. For polyclonal stimulation, the recommended cell concentration per well should be reduced to  $2 \times 10^2$  -  $10^5$  cells per well.

### **NOTES:**

- It is recommended to test the samples in triplicate and in serial dilutions in the ELISPOT procedure.
- No more than  $3 \times 10^5$  cells/well should be suspended in the ELISPOT plate. At higher concentrations the cells will be stacked upon each other, resulting in poor spot formation.

See also 'Guidelines for cell concentrations and cell incubation times' on page 17 for more information on cell concentration/well, incubation times and stimuli.

# ELISPOT procedure

*All solutions should be at RT prior to use. The steps 1 till 9 should be performed under sterile conditions. In addition, estimate the time needed to prepare all cell preparations which have to be ready for step 7 and plan accordingly.*

1. Add 50  $\mu$ l of diluted coating antibody solution into each well of the ELISPOT plate and fill up to 100  $\mu$ l with PBS-I.
2. Cover the plate with a lid and incubate overnight at 4°C.
3. Remove coating antibody solution and rinse each well 3x with 200  $\mu$ l PBS-I. The plate is subsequently emptied.
4. Add 200  $\mu$ l Blocking buffer (1x) into each well.
5. Cover the plate with a lid and incubate for at least 1 hour at RT. During this incubation step start preparing the cell sample preparations (see “Cell sample preparation” on page 14).
6. If the cell preparations are ready, decant the blocking buffer from wells (do not wash the wells).
7. Bring the cell preparation into the wells of the ELISPOT plate. Add 100  $\mu$ l/well.
8. Cover ELISPOT plate with lid and incubate at 37°C, 5% CO<sub>2</sub> and 100% humidity. The incubation time can vary from 24 to 72 h. Specific activation conditions will vary, depending on cell type, signaling protein of interest, kinetics of protein release and whether a preincubation step was included in the procedure. See “Cell sample preparation” on page 14, “Guidelines for cell concentrations and cell incubation times” on page 17 and Typical data.
9. Remove the bulk of cells with a firm shake-out action and rinse each well 2x with 200  $\mu$ l PBS-I. The plate is subsequently emptied.
10. Wash the plate 6x with 250  $\mu$ l Wash buffer/well (see “Addendum T cell ELISPOT assay” for directions on washing).
11. Add 100  $\mu$ l of diluted biotinylated detection antibody into each well.
12. Seal the plate with adhesive cover slip and incubate 1 hour at 37°C (or overnight at 4°C).
13. Empty plate and wash 6x with Wash buffer.
14. Add 50  $\mu$ l diluted GABA conjugate into each well.
15. Seal the plate with an adhesive cover slip and incubate 1 hour at 37°C.
16. Empty plate and wash 6x with Wash buffer and subsequently empty by a firm shake (wells should not contain residual Wash buffer).

Sterile conditions

Non sterile conditions



17. Add 35 µl freshly prepared Activator I/II solution into each well. Distribute the Activator I/II uniformly over the bottom of the well.
18. Cover plate with lid and incubate for 30-60 min at RT. Monitor spot development by light microscope.
19. When clear spots have developed, stop the reaction by emptying the plate and rinse with demineralized water.
20. Air-dry the plate at RT.
21. Count spots by use of a transmitted light microscope or an Immunospot image analyzer.

*Visually, spots have a grayish color, microscopically they are black. Silver-stained spots are highly stable and spot quality is preserved indefinitely when the plate is stored at a dry place.*

## Guidelines for cell concentrations and cell incubation times

Cell type	Analyte <sup>stimuli</sup>	Incubation time without preincubation	Incubation time after preincubation*	Cell concentration antigen stimulation	Cell concentration polyclonal stimulation
Human and Old world Monkey PBMCs	IFN-γ <sup>1-4,6,7</sup> IIL-12p70 <sup>**5</sup> IL-2 <sup>1-4,6,7</sup> IL-12/23p40 <sup>5</sup> IL-4 <sup>1-4,6,7</sup> IL-13 <sup>1-4,7</sup> IL-5 <sup>1-4,7</sup> IL-17A <sup>1-4,7</sup> IL-6 <sup>1-3,5</sup> IL-17F <sup>1</sup> IL-10 <sup>1-5,7</sup> TNF-α <sup>1-3</sup>	24-48h	24h	2x10 <sup>5</sup> cells/well	2x10 <sup>3</sup> to 1x10 <sup>5</sup> cells/well
	Perforin <sup>1,2,6</sup>	48-72h	24h	2x10 <sup>5</sup> cells/well	2x10 <sup>3</sup> to 1x10 <sup>5</sup> cells/well
Rodent spleen cells	IFN-γ <sup>1-3</sup> IL-6 <sup>1-3</sup> IL-2 <sup>1-3</sup> IL-10 <sup>1-3</sup> IL-4 <sup>1-3</sup> TNF-α <sup>1-3</sup> IL-5 <sup>1-3</sup>	24-48h	24-48h	2x10 <sup>5</sup> cells/well	2x10 <sup>2</sup> to 1x10 <sup>5</sup> cells/well

**General cell stimuli commonly used in ELISPOT**

Polyclonal stimuli: 1. PMA & ionomycin (50 ng/ml & 1 µg/ml) 2. PHA (10-30 µg/ml) 3. ConA (human and Old world monkey: 6-10 µg/ml) (rodent: 4 µg/ml) 4. α-CD3 & α-CD28 (0.05 µg/ml & 0.05 µg/ml) 5. LPS & IFN-γ (100 ng/ml & 10-100 units/ml)

Antigen-specific stimuli (human PBMCs): 6. ICE peptide pool (1 µg/ml for each peptide) 7. Tetanus toxoid (0.5 LF/ml)

\*Preincubation time: 24-48h (all cell types); cell concentration 4x10<sup>6</sup> cells/ml

\*\*NOTE: Preincubation is not recommended for IL-12p70.

# Notes

# Notes

# Technical assistance

If you require assistance, information or have any questions, please contact our company:

**U-CyTech biosciences**

Phone: +31.30.253 5960

E-mail: [info@ucytech.com](mailto:info@ucytech.com)

On our website ([www.ucytech.com/manuals](http://www.ucytech.com/manuals)) you can find: Manuals, Typical data, Addendum and MSDS of our ELISPOT kits.

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